

(b) The half-contraction time (CT): This was taken as the time after start of stimulation for force to reach 50% maximum force during a test contraction.

(c) The rate of relaxation₅₀₋₃₀ (RT): This was taken as the time taken for the force of contraction to decline from 50% to 30% of the peak value during the test contractions.

(d) Fatigue resistance (ratio of the peak tetanic contraction before and after the stimulation protocol).

The experiment was conducted in two groups: the control (Group-A) and the test groups (B & C). In the test group, Aminophylline was added to the Ringer solution bathing the nerve-muscle preparation. Two strengths of Aminophylline was used (a) 50 µM/L (Group-A) and (b) 500 µM/L (Group-B). Fatiguability was tested after allowing a 30-minute equilibrium period (7).

RESULTS

The peak contraction, half-contraction time and relaxation₅₀₋₃₀ time for the different groups

at rest and after fatiguing stimulation are given in Tables Ia and Ib. Aminophylline did not appear to have any effect on the peak contraction, half-contraction time and relaxation₅₀₋₃₀ time of the resting muscle as analysed by Wilcoxon Matched Pair Signed Rank Test. However, aminophylline did appear to affect some if not all of the post-fatigue changes. The Kruskal-Wallis' Test and Mann-Whitney U Test showed that the increase in the half-contraction and relaxation₅₀₋₃₀ periods following muscle fatigue were susceptible to the effects of aminophylline (Table II). In general, the results of our study indicate that aminophylline in high concentration (500 µM/L) minimises the increase in half-contraction period of isolated frog gastrocnemius muscle preparations in response to a high-energy-demand stimulation protocol of fatigue-induction. At the same time, it prolongs the increase in relaxation₅₀₋₃₀ time in response to the same protocol. Aminophylline did not affect the fatigue-induced changes in the peak tetanic contraction, the half contraction time or the relaxation₅₀₋₃₀ time.

TABLE I (a): The mean \pm SD values of peak contraction, half-contraction time and relaxation₅₀₋₃₀ time in Groups A, B and C, measured before a fatiguing stimulation, and after fatiguing stimulation protocol 1.

Parameter	Group-A	Group-B	Group-C
1. Peak contraction in control		0.57 \pm 0.13	0.80 \pm 0.16
2. Half-contraction time in control		0.05 \pm 0.02	0.05 \pm 0.01
3. Relaxation ₅₀₋₃₀ time in control		0.07 \pm 0.04	0.05 \pm 0.02
4. Peak contraction before protocol 1	0.58 \pm 0.04	0.62 \pm 0.12	0.81 \pm 0.16
5. Half-contraction time before protocol 1	0.04 \pm 0.01	0.05 \pm 0.02	0.05 \pm 0.01
6. Relaxation ₅₀₋₃₀ time before protocol 1	0.06 \pm 0.01	0.06 \pm 0.05	0.05 \pm 0.02
7. Fatigue resistance in protocol 1	48.30 \pm 19.40	48.40 \pm 16.20	0.60 \pm 4.09
8. Peak contraction after protocol 1	0.25 \pm 0.08	0.34 \pm 0.20	0.33 \pm 0.16
9. Half-contraction time after protocol 1	0.12 \pm 0.03	0.12 \pm 0.03	0.08 \pm 0.01
10. Relaxation ₅₀₋₃₀ time after protocol 1	0.06 \pm 0.01	0.07 \pm 0.04	0.12 \pm 0.01

TABLE I (b): The mean \pm SD values of peak contraction, half-contraction time and relaxation₅₀₋₃₀ time in Groups A, B and C, measured before a fatiguing stimulation, and after fatiguing stimulation protocol 2.

<i>Parameter</i>	<i>Group-A</i>	<i>Group-B</i>	<i>Group-C</i>
1. Peak contraction in control		0.57 \pm 0.13	0.80 \pm 0.16
2. Half-contraction time in control		0.05 \pm 0.02	0.05 \pm 0.01
3. Relaxation ₅₀₋₃₀ time in control		0.07 \pm 0.04	0.05 \pm 0.02
4. Peak contraction before protocol 2	0.50 \pm 0.03	0.57 \pm 0.07	0.73 \pm 0.16
5. Half-contraction time before portocol 2	0.06 \pm 0.01	0.05 \pm 0.01	0.06 \pm 0.02
6. Relaxation ₅₀₋₃₀ time before protocol 2	0.07 \pm 0.01	0.11 \pm 0.03	0.11 \pm 0.01
7. Fatigue resistance in protocol 2	10.40 \pm 1.50	8.80 \pm 4.10	7.96 \pm 1.58
8. Peak contraction after protocol 2	0.33 \pm 0.05	0.34 \pm 0.09	0.45 \pm 0.22
9. Half-contraction time after protocol 2	0.10 \pm 0.01	0.12 \pm 0.04	0.14 \pm 0.02
10. Relaxation ₅₀₋₃₀ time after protocol 2	0.09 \pm 0.01	0.16 \pm 0.02	0.11 \pm 0.01

TABLE II : Comparison of fatigue-induced changes (i.e. changes in peak contraction, half-contraction time and relaxation₅₀₋₃₀ time) in Group A, B and C by the Kruskal-Wallis' Test. Significance of multiple comparison is by Mann-Whitney 'U' Test where comparison is significant.

<i>Indices of Muscle Fatigue</i>	<i>Inter-group comparisons</i> <i>Significance at P = 0.05</i>
Fatigue resistance in protocol 1	Not significant
Fatigue resistance in protocol 2	Not significant
Change in peak contraction after protocol 1	Not significant
Change in half-contraction time after protocol 1	Group A & C are significantly different (P<0.001) Group A & B are not significantly different
Change in relaxation ₅₀₋₃₀ time after protocol 1	Group A & C are not significantly different (P<0.001) Group A & C are significantly different
Change in Half-contraction time after protocol	Not significant
Change in relaxation ₅₀₋₃₀ time after protocol 2	Not significant
Change in relaxation ₅₀₋₃₀ time after protocol 2	Not significant

DISCUSSION

Aminophylline is known to increase the twitch and submaximal tetanic force generation of mammalian diaphragm (1, 2) and frog semitendinosus muscle (3). Aminophylline is known not to affect maximum tetanic force of mammalian (2) or frog muscle (3).

In our study, we have used two protocols of fatigue stimulation, but although both of them were tetanic stimulations, both caused submaximal contractions (The stimulation frequencies were 100Hz and 30Hz).

Some of the effects of aminophylline can be inhibited by removing extra-cellular calcium or

by blocking the influx of calcium through the plasma membranes (1, 2). Several groups have proposed that aminophylline increases the force at submaximal stimulation by increasing calcium influx across the sarcolemma during excitation-contraction coupling (1, 2, 8, 9). In addition, at submaximum frequencies of stimulation, increased calcium influx may increase calcium release from the sarcoplasmic reticulum by a calcium-dependent calcium release mechanism (9).

The potentiation of submaximum force development by aminophylline has been shown

to be inhibited by calcium channel blockers (1). Consequently, aminophylline appears to enhance the force development of submaximally activated muscle by increasing intracellular calcium (11) and activation additional crossbridges (2, 3).

It is possible that aminophylline minimises the fatigue-induced increase in contraction time by increasing the entry of Ca^{++} into the sarcoplasm. The prolongation of fatigue-induced increase in relaxation time can also be attributed to the increased Ca^{++} entry.

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